

Implementation of SNOMED CT in Histopathology and Genomics for Cancer Care

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Nebraska
Medicine



Begin with the end in mind

- Render pathology data to computable forms for patient care at the point of care
- Render genomics data to computable form for use at the point of care for patient care
- Capture data in pathology and genomics at the point of care to support new discovery at the bench
- Bring bench discovery back to the point of care to support patient care

**It is about the patient and aiding
the patient and care team to
make informed decisions**





Multiple Use Cases

Cancer treatment planning

Precision medicine research project planning

Laboratory risk and safety management

Retrieving biobank tissue for research protocols



Tangible Examples

Cancer treatment planning:

- Colon cancer with BRAF V600E mutation and KRAS mutation in codon 12. Anti-EGFR therapy contraindicated.

Research Project Planning:

- How many healthy patients do we have that are BRCA1 or BRCA2 positive
- Retrieve all breast cancer cases that tested ER-, PR- and Her2/neu

Medical-legal, compliance and safety:

- Recall of all cancer cases for treatment review with (formerly thought insignificant) somatic gene sequence variant reported
- Accreditation requirements for molecular laboratories.

Tissue Biobank Applications:

- Find all malignant neoplasms of any origin tested for BRAF mutation AND reported positive for lymphatic metastases.





Terminology Implementation Requirements

- Conform to the current EHR ecosystem
- Incorporation into natural workflows of providers
 - Upstream systems
 - Point of care
 - Non-intrusive
- Support primary and secondary uses of data





EHR Technology

- What it is:
 - Patient-centric
 - Encounter based
 - Longitudinal Medical Record
 - Transactional
 - Billing orientation
- What it can be:
 - Integrated patient-centric data
 - Basis for clinical decision support
 - Population management
- Requirements (Just a few...)
 - Standards
 - Standard use of standards
 - System Integration/Interoperability



Histopathology Reporting Evolution

Basic

Advanced



Level 1 -
Narrative

Level 2 -
Narrative
with
required
data
elements

Level 3 -
Narrative
with
required
data
elements
in
Synoptic
format

Level 4 -
Level 3
plus
electronic
user
interface
for data
entry

Level 5 -
Level 4
plus
structured
language
and
discrete
data
capture

Level 6 -
Level 5 plus
all data
encoded in
machine
readable,
standard
terminology

Level 7-
Semantic
interoperabi
lity

1. Srigley JR, McGowan T, Maclean A, Raby M, Ross J, Kramer S, et al. Standardized synoptic cancer pathology reporting: a population-based approach. J Surg Oncol. 2009 Jun 15;99(8):517-24.

Sample Pathology Reports (1990 – 2005); Level <3

I: Pancreatic resection with adherent duodenal resorption (measuring 11 cm), ventricular resorption (measuring approximately 5 cm), and gall bladder, macroscopic u a. **Pancreatic resection** approximately 8 x 4 cm, with a **3 x 3 cm tumor**, cut and chopped and constricted choledochus and pancreatic cancer.

Histologically, the corresponding tumor, infiltratively growing atypical gland formations, is seen. Cylindrical gland epithelium with nuclear stratification, enlarged hyperchromatic cell nuclei and mitosis. Central tumor necrosis. Picture as in **medium differentiated adenocarcinoma**. Macroscopically and with taken histological cuts, **radically excised with narrow but free margin against the retinal vena porta.** (B). The medi preparation contains a **reactive lymph node**.

I (1): 7.7 cm ventricle with ...until the papilla. Continuous 8.5 x 2.5 cm gallbladder. 2.5 x 3 x 2 cm **yellow-white tumor-like change, growing partially in the pancreatic head against the duodenal mucosa and choledochus and papilla vateri.**

The ventricular free ducts, duodenum, leftover pancreatic tissue and choledochus without detectable tumor growth. In the gall bladder, microscopic focal hyperplastic mucosa and signs of chronic cholecystitis with lymphocytic infiltration are observed. No signs of malignancy in the gall bladder.

Similar to the macroscopic tumor, a **medium to focal low differentiated adenocarcinoma** is seen that **grows under the duodenal mucosa and into the pancreatic head** with large necrotic areas and desmoplastic connective tissue formation. ...comprised of major tubular formations, means that **one should primarily suspect the outcome of proximal choledochus or pancreatic cancer.** Biggest tumor size 2.5 x 2.2 cm. Distance to the nearest travel area 1.6 cm.

In fraction I, 21 tumor-free lymph nodes are found. In addition, two tumor tumors of tumor growth per continuitatem from the tumor.
T3 NO MX.



Sample Pathology Report – 2017 (Level 4)

Type of preparation:

- Whipple

Microscopic assessment

Origin: **Pancreas**

Histological Type: Ductal Adenocarcinoma, with partial foamy gland pattern

Differential rate: well to moderate

Corrected tumor size:

- Craniocaudal: 3.1 cm (Slices 2 to 8)
- Axial: 3.7 x 2.2 cm (in large section Z / disc 6).

Tumor growth in neighboring organs / structures:

- The major part of the tumor grows in the cranial and central regions of the pancreatic head.
- Tumor invades the peripancreatic fat tissue, the bile duct and extensively the duodenal wall, focally up to the mucosa.

Tumor shows intensive vascular invasion / spread as well in venous and lymphatic vessels.

Lymph vessel growth: extensive invasion present

Vascular invasion: extensive invasion present in multiple medium-sized veins, partially with intraluminal tumor and partially obliterated.

Perineural invasion: present

Distance from tumor to nearest area of travel:

- Tumor cells present focally <1 mm from cranial (lig. Hepatoduodenal) and posterior margins

Regional lymph nodes:

With metastasis: 5 (including 1 from complementary preparations T 794-17)

Total: 17 (including 1 from Supplementary Preparations T 794-17)

subfractionation:

- 0/3 inferiora
- 1/4 anteriora
- 0/1 against SMIEs
- 2/2 against SMA
- 1/1 periductala
- 0/3 oment
- 1/2 station 8A (from preparation T794-17)

The major part of the tumor grows in the cranial and central regions of the pancreatic head.

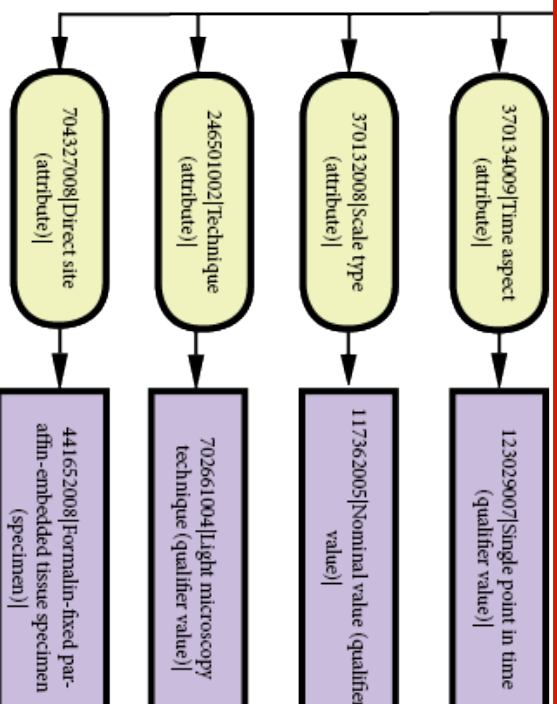
- With metastasis: 0



CAP Approved

Microscopic Tumor Extension

- Cannot be assessed
- No evidence of primary tumor
- No invasion (high-grade dysplasia/intraepithelial carcinoma)
- Tumor invades lamina propria/muscularis mucosae (intramucosal)
- Tumor invades submucosa
- Tumor invades muscularis propria
- Tumor invades through the muscularis propria into the subserosal pericolic or perirectal soft tissues but does not extend to the serosa
- Tumor penetrates to the surface of the visceral peritoneum (serosa)
- Tumor is adherent to other organs or structures (specify: _____)
- Tumor directly invades adjacent structures (specify: _____)
- Tumor penetrates to the surface of the visceral peritoneum (serosa) (specify: _____)



AP
Example:
Microscopic
local
invasion of
colon tumor



Example Value set

Direct extension of colon tumor	785953131000004104 Status of microscopic invasion of excised colon neoplasm (observable entity)
Tumor invasion cannot be assessed	87100004 Topography unknown (body structure)
No evidence of primary tumor	21229009 Topography not assigned (body structure)
Carcinoma in situ, intraepithelial	42978003 Colonic epithelium (body structure)
Carcinoma in situ, invasion of lamina propria	113284008 Colonic lamina propria (body structure)
Tumor invades submucosa	61647009 Colonic submucosa (body structure)
Tumor invades muscularis propria	41948009 Colonic muscularis propria structure (body structure)
Tumor invades through the muscularis propria into the subserosa or into non-peritonealized pericolic or perirectal tissue	52010009 Colonic subserosa (body structure)
Tumor penetrates serosa	90132000 Colonic serosa (body structure)

Value set of answers



Implementation - Terms Bound to CoPath® for Pathologist

COLON AND RECTUM: Resection

Page 2 of 8

Tumor Size	
F1 Greatest dimension: 2.2 cm	
F2 * Additional dimensions: _____ cm	
F3 Cannot be determined	
F4 Other (specify): _____	
Macroscopic Tumor Perforation	
G1 Present	
G2 Not identified	
G3 Cannot be determined	
* Macroscopic Intactness of Mesorectum	
H1 * Not applicable	
H2 * Complete	
H3 * Near complete	
H4 * Incomplete	
H5 * Can not be determined	
H6 * Other (specify): _____	
**** NOTE ****	
All rectal carcinomas arising distal to peritoneal reflection, should have notation regarding mesorectum.	
Histologic Type	
J1 Adenocarcinoma	
J2 Mucinous adenocarcinoma (greater than 50% mucinous)	
J3 Signet-ring cell carcinoma (greater than 50% signet-ring cells)	
J4 High-grade neuroendocrine carcinoma	
J5 Large cell neuroendocrine carcinoma	
J6 Small cell neuroendocrine carcinoma	
J7 Squamous cell carcinoma	
J8 Adenosquamous carcinoma	
J9 Medullary carcinoma	
J10 Undifferentiated carcinoma	
J11 Other (specify): _____	
J12 Carcinoma, type cannot be determined	
Histologic Grade	
K1 Not applicable	
K2 Cannot be determined	
K3 Low-grade (well to moderately differentiated)	
K4 High-grade (poorly differentiated to undifferentiated)	
K5 Other (specify): _____	



Resultant Report

MICROSCOPIC TUMOR CHARACTERISTICS

Histologic type of neoplasm:	Mucinous adenocarcinoma (greater than 50% mucinous)
Histologic grade of neoplasm:	Low-grade (well to moderately differentiated)
Mucinous histologic fraction of neoplasm:	(%): 95
Percent signet ring cells in adenocarcinoma:	(%): 0
Intratumoral Lymphocytic Response:	None
Peritumoral Lymphocytic Response:	None
Status of tumor budding in carcinoma:	None
Number of tumor buds per HPF (Average per 10 HPF):	Average # per HPF: 0
Perineural Invasion:	Perineural invasion absent
Lymphatic (Small Vessel) Invasion (L):	Absent
Intramural vascular (Large vessel) invasion:	Absent
Extramural vascular (Large vessel) invasion:	Absent
Polyp Type in which invasive carcinoma arose:	None identified

ANCILLARY TESTING

Mismatch repair abnormality by IHC:

MLH1 - Mismatch Repair (MMR) Proteins by IHC:	No: Mismatch repair proficient
MSH2-Mismatch Repair (MMR) Proteins by IHC:	Intact nuclear expression
MSH6-Mismatch Repair (MMR) Proteins by IHC:	Intact nuclear expression
PMS2-Mismatch Repair (MMR) Proteins by IHC:	Intact nuclear expression
BRAF Expression (by immunohistochemistry):	Negative for cytoplasmic expression



CoPath® Structured AP Synoptic Reports

```
MSH|^~\&|COPATH|COPATH|EPIC||2017NNNN||ORU^R01||P|2.3.1
PID|1||2529400||NAME^^^|1968NNNN|M||W|ADDRESS||
PV1|1||3BE|||||||||^NAME|TCE
OBR|1||SNN-NNNN^CoPathPlus|EXAM^EXAM|||||A|^RIGHT COLON|NNNNNN|||||F
OBX|2|CWE|721754191000004103^Colon-Margins: Distal^SCT||369708000^Distal margin
uninvolved by invasive carcinoma^SCT|||||F
OBX|3|CWE|200661231000004105^Colon-Specimen(s) included in case^SCT||
32713005^Cecum^SCT|||||F
OBX|4|CWE|661259881000004105^Cancer Treatment Effect^SCT||997170731000004106^No
prior treatment^SCT|||||F
OBX|5|CWE|669509521000004104^Cancer Perineural Invasion^SCT||370051000^Perineural
invasion absent^SCT|||||F
OBX|6|CWE|384625004^Primary Tumor (pT) Category^SCT||395707006^pT3: Tumor invades
through the muscularis propria into pericolorectal tissues^SCT|||||F
OBX|7|CWE|371497001^Colon-Distant Metastasis (pM)^SCT||17076002^PMX: Cannot be
assessed^SCT|||||F
OBX|9|CWE|200661231000004105^Colon-Specimen(s) included in case^SCT||
66754008^Appendix^SCT|||||F
OBX|10|CWE|200661231000004105^Colon-Specimen(s) included in case^SCT||
9040008^Ascending colon^SCT|||||F
```

EHR Technology - Genomics

- What it is:
 - Patient-centric
 - Encounter based
 - Longitudinal Medical Record
 - Transactional
 - Billing orientation
- What it can be:
 - Integrated patient-centric data
 - Basis for clinical decision support
 - Population management
- Requirements (Just a few...)
 - Standards
 - Standard use of standards
 - System Integration/Interoperability



Houston: We have a problem



How to put a square peg into a round hole?

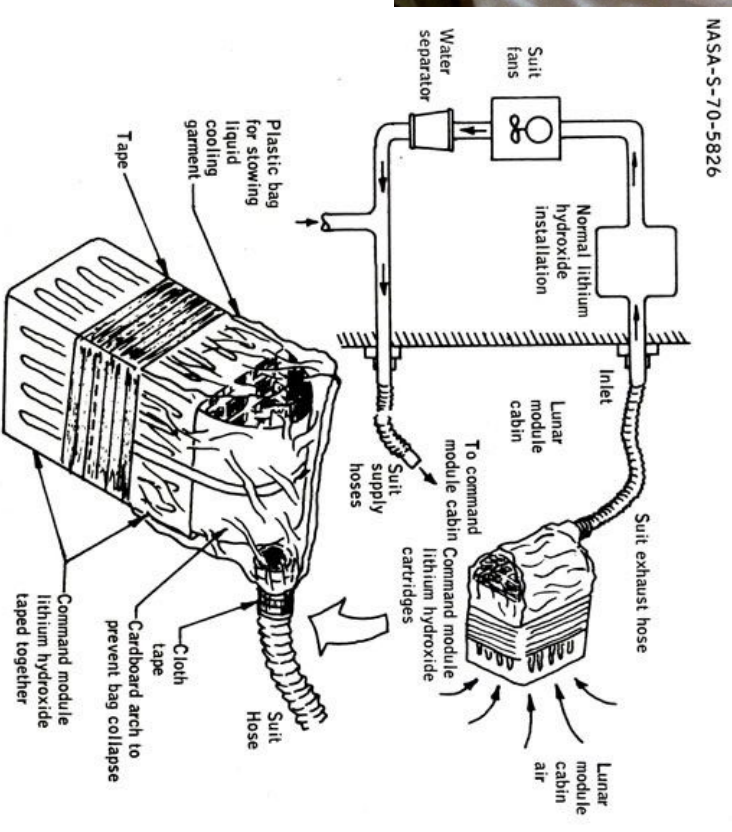


Figure 6.7-1.- Supplemental carbon dioxide removal system.

What to do with all this Data?

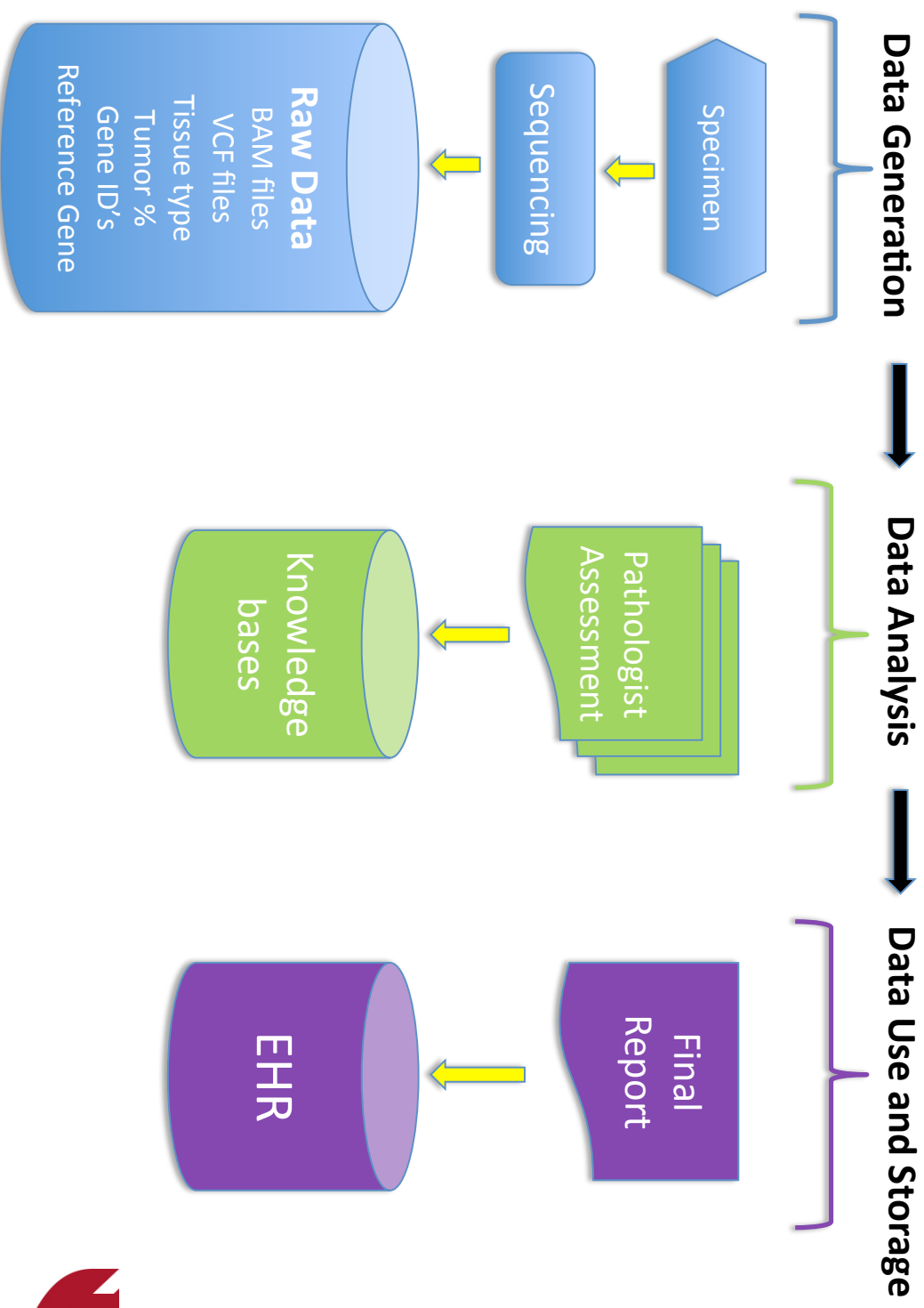
Technical Desiderata for integration of genomic data into the EHR

1. Maintain separation of primary molecular observations from the clinical interpretations of those data
2. Support lossless compression from primary molecular observations to clinically manageable subsets
3. Maintain linkage of molecular observations to the laboratory methods used to generate them
4. Support compact representation of clinically actionable subsets for optimal performance
5. Simultaneously support human viewable formats and machine readable formats in order to facilitate implementation of decision support rules
6. Anticipate fundamental changes in the understanding of human molecular variation
7. Support both individual clinical care and discovery science

1. Masys DR, Jarvik GP, Abernethy NF, Anderson NR, Papanicolaou GJ, Pattoo DN, et al. Technical desiderata for the integration of genomic data into Electronic Health Records. *J Biomed Inform.* 2012 Jun;45(3):419-22.

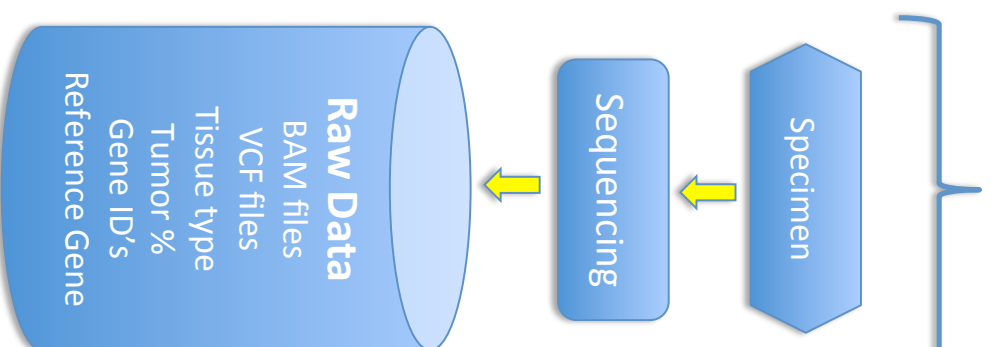


Molecular Pathology Data Flow



Data Use – Data Generation

Data Generation



Raw Data (i.e., Primary Data)

Necessary for clinical interpretation

Valuable to Research Community

Value to consumers – clinicians,
patients?



Data Use – Data Analysis

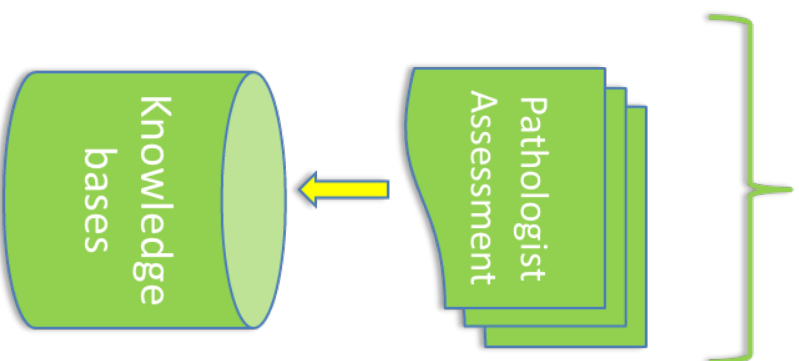
Data Analysis

Pathologists consider outputs of Data Generation step

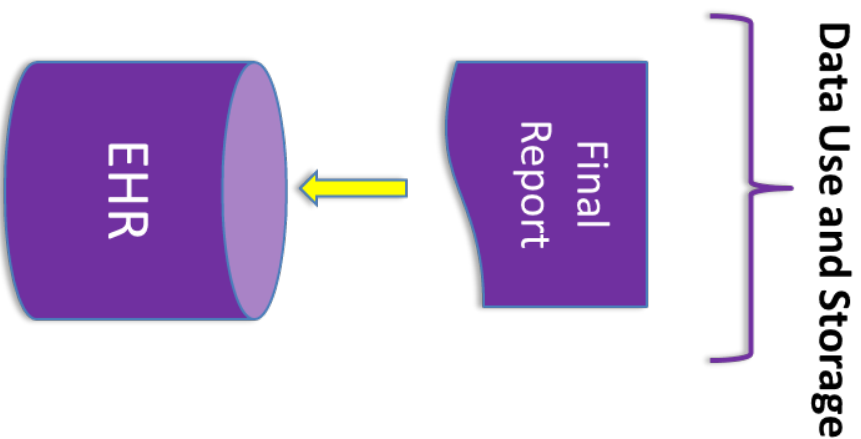
Access Knowledge Bases to reconcile data with current genomic understanding

Render assessment of variants detected and should be reported

Classify variants in terms of clinical significance



Data Use – Use and Storage



Data Use and Storage

Final report – pdf format

Enumeration of variants detected
classified by clinical significance

Additional items: Type of alteration,
Allele frequency (in population),
availability of potential clinical trials

Useful for individual clinical
encounter but of limited value for
extended clinical care, decision
making and research.





More on the Final Report

- Based on the specific gene variants detected and **reported** by the pathologist.
- What elements are needed in the EHR in computable form?
- One-to-one relationship of reported variants and their representation in the variant call file (VCF)
- Can the VCF representation of the reported variants be used as discrete elements in the EHR?



Variant Call Format

```
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT NORMAL TUMOR
20 14370 rs6054257 G A 29 PASS NS=3;DP=14;AF=0.5;DB;H2 GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:3:5:65,3
20 17330 . T A 3 q10 NS=3;DP=11;AF=0.017 GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:3:5:65,3
20 1110696 rs6040355 A G,T 67 PASS NS=2;DP=10;AF=0.333,0.667;DB GT:GQ:DP:HQ 1|2:21:6:23,27 2|1:2:0:18,2
20 1230237 . T 47 PASS NS=3;DP=13;AA=T GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51
20 1234567 microsat1 GTC G,GTCTC 50 PASS NS=3;DP=9;AA=G GT:GQ:DP 0/1:35:4 0/2:17:2
```

Important data

Essential data for EHR?

Discrete and computable

Contains data that has limited value outside the context of the initial report

Difficult to import and manage within EHR structures

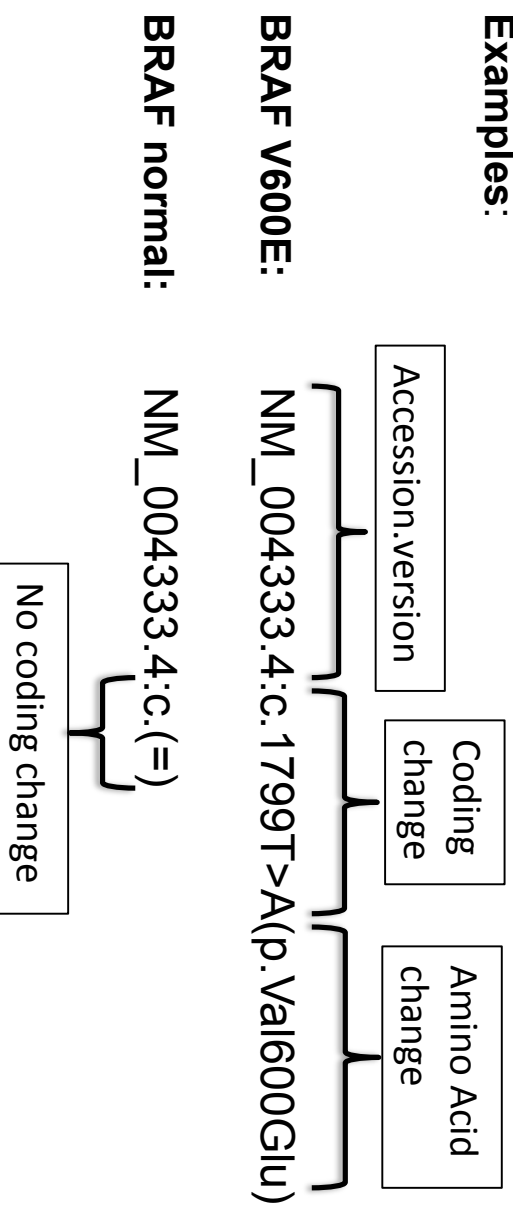
TOO COMPLICATED FOR EHR – Need something simple



Human Genome Variation Society Representation

- Human Genome Variation Society (HGVS)
- Standardized nomenclature for variants
- Includes nomenclature for normal sequences

Examples:



HGVs in EHR and Biobank

- HGVs representation of genetic information in EHR is tractable
- Data is a structured string
 - Easily accommodated in EHR
 - Data easily queried by regular expression
- Maintain links to curated gene knowledge bases
 - Standardized representation
 - Used by reference databases, knowledge bases
- Applicable to targeted gene sequencing and whole genome sequencing
- Retains clinically relevant and actionable information
- Can be readily moved via HL7



Moving Molecular Data

HL7 Lab order and results message types, of course

Orders:

Specimen Type – SNOMED CT

Ordered Service – LOINC

Diagnosis – ICD-10 or SNOMED CT

Results:

Performed tests – LOINC (OBR-4)

Results – LOINC (OBX-3) and HGVS (OBX-5)

Abnormal flag – Association of Molecular Pathologists Tiers 1 - 4

Example:

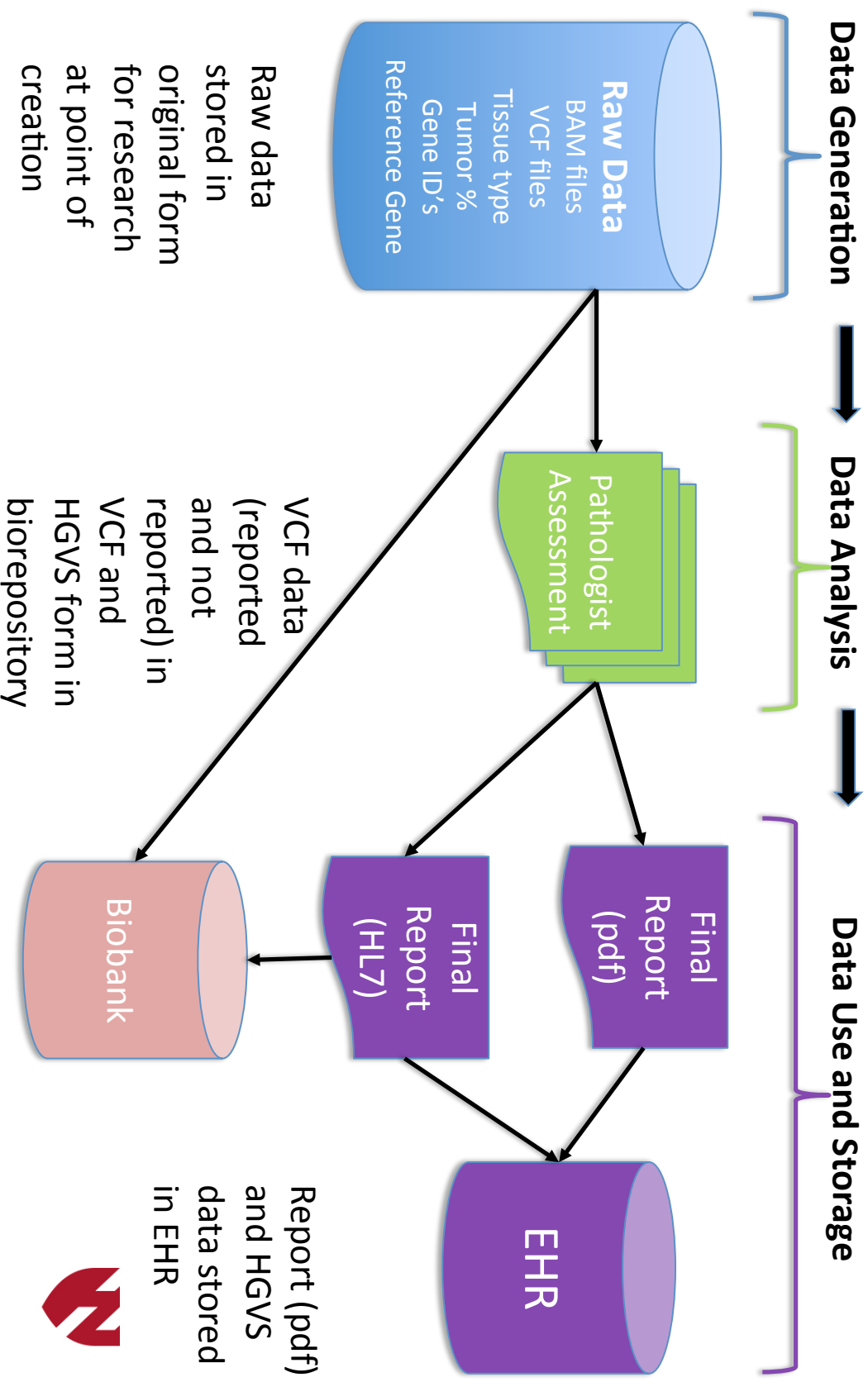
OBR|1|CE|segment 3|51966-0^Genetic disease DNA analysis panel^LN|....

OBX|1|CE|85511-4^BRAF gene mutations found in colorectal cancer specimen by molecular genetics method^LN||NM_004333.4(BRAF):c.1799T>A (p.Val600Glu)|...| Pathogenic|

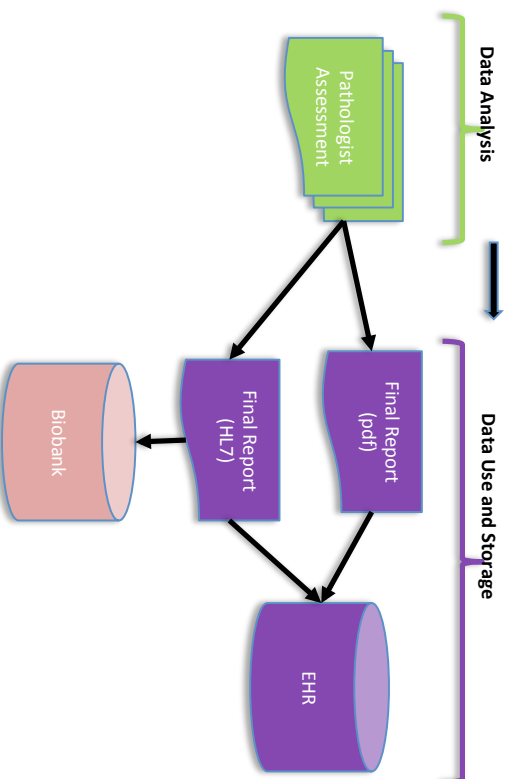
OBX|2|CE|85509-8^KRAS gene mutations found in colorectal cancer specimen by molecular genetics method^LN||NM_004985.4(KRAS):c.35G>A (p.Gly600Asp)|...| Pathogenic|



Genomic Data Flow and Store



Genomic Data Flow Example



1. Pathologist sign-out is the trigger event
2. PDF report sent per usual practice
3. HL7 version 2.5.1 message sent to biobank and EHR, simultaneously

HL7 Message Sent

```

MSH|^~\&|GenomOncology|Workbench|UNMCM|irth|UNMCM||ORU^R01^ORU_R01|77801.1|P|2.5.1|
PID|1||12345||Doe^Jane^||19850206|F
ORC|1||G17-xxx||CM||^v^v^
OBR|1||G17-xxx|55232.3^Genetic analysis summary panel^LN|||
OBR|1|FT|51969.4^Genetic analysis summary report^LN||<P>For this specific specimen there was 200X coverage for the following regions,
therefore low frequency variants in these regions may not be identified: three amplicons of CEBPA exon1, CUX1 exons 1, 19, and 23, and STRA2 exon 7.
Only clinical trials that pertain to genes with identified somatic mutations are reported.
OBR|2||G17-xxx|55207.5^Genetic analysis discrete result panel^LN|||
OBR|1|CWE|911752541000004109^TP53 sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|TP53 NP_000537.3:R175H NM_000546.5:c.
524G>A^TP53 R175H|||Pathogenic|||F
OBR|2|CWE|911752871000004102^ASXL1 sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|ASXL1 NP_056153.2:N9865 NM_015338.5:c.
2957A>G^ASXL1 N986S|||Likely Benign|||F
OBR|3|CWE|911752061000004102^ABL1 sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|ABL1 NM_005157.4:c.(=)|||Normal|||F
OBR|4|CWE|911752881000004104^ATRX sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|ATRX NM_000489.3:c.(=)|||Normal|||F
OBR|5|CWE|911752891000004101^BCOR sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|BCOR NM_001123385.1:c.(=)|||Normal|||F
OBR|6|CWE|911752901000004102^BCORL1 sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|BCORL1 NM_021946.4:c.(=)|||Normal|||F
OBR|7|CWE|911752111000004101^BRAF sequence variant identified in excised malignant neoplasm (observable entity)^SCT|1|BRAF NM_004333.4:c.(=)|||Normal|||F

```



EPIC Clinician View

Genetic analysis discrete result Visible to patient: No (Not Released) Next appt: None
 Collected: 9/20/2017 16:42 Status: Final result 16:42

Genetic analysis summary report Here are my 2nd comments:
 KRAS seq. variant ID'ed in excised malignant KRAS NP_004976.2:Q61H NM_004985.3:c.183A (Pathogenic)
 neoplasm
 KRAS seq. variant ID'ed in excised malignant KRAS NP_004976.2:Q61Y NM_004985.3:c.181_183delCAalnSTAC (Likely Patho)
 neoplasm
 Observable Entity AKT1 NM_001014432.1:c(=)
 Comments: Normal
 BRAF seq. variant ID'ed in excised malignant BRAF NM_004333.4:c(=)
 neoplasm
 Comments: Normal
 EGFR seq. variant ID'ed in excised malignant EGFR NM_005228.3:c(=)
 neoplasm
 Comments: Normal
 ERBB2 seq. variant ID'ed in excised ERBB2 NM_004448.2:c(=)
 malignant neoplasm
 Comments: Normal
 ERBB4 seq. variant ID'ed in excised ERBB4 NM_005235.2:c(=)
 malignant neoplasm
 Comments: Normal
 NRAS seq. variant ID'ed in excised malignant NRAS NM_002524.4:c(=)
 neoplasm
 Comments: Normal
 PIK3CA seq. variant ID'ed in excised PIK3CA NM_006218.2:c(=)
 malignant neoplasm
 Comments: Normal
 PTEN seq. variant ID'ed in excised malignant PTEN NM_000314.4:c(=)
 neoplasm
 Comments: Normal
 Resulting Agency/ COPATH

Narrative
 Case number: Integration 3- Lung
 Specimen Collected: 09/20/17 16:42 Last Resulted: 09/20/17 16:57 [Lab Flowsheet](#)

Specimen Source
 Lung

© Epic Computer Systems. Used with permission.

© Nebraska Lexicon (SNOMED CT extension)



Patient Care Use

Find all patients with colorectal cancer AND a mutation in the PIK3CA locus and/or the ERBB2 gene locus.

Aspirin therapy shown to be beneficial

DeidPatId	Observable	HGVs	Pathogenicity
"100020"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NP_006209:C420R NM_006218:c.1258T>C"	"Uncertain Significance"
"100020"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NP_006209:E453del NM_006218:c.1359_1361delAGA"	"Likely Pathogenic"
"100020"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100008"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NM_006218.2:c.(=)"	"Normal"
"100008"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100285"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA null:E545K null:c.1633G>A"	"Pathogenic"
"100195"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NP_006209:R91M NM_006218:c.1173A>G"	"Likely Benign"
"100195"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NP_006209:E109_I112delinsD NM_006218:c.327_335delAGAAAAGAT"	"Likely Pathogenic"
"100195"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100061"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NM_006218.2:c.(=)"	"Normal"
"100061"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100063"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NP_006209:R91M NM_006218:c.1173A>G"	"Likely Benign"
"100112"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100112"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA NM_006218.2:c.(=)"	"Normal"
"100249"	"ERBB2 sequence variant identified in excised malignant neoplasm (observable entity)"	"ERBB2 NM_004448.2:c.(=)"	"Normal"
"100249"	"PIK3CA sequence variant identified in excised malignant neoplasm (observable entity)"	"PIK3CA null:E542K null:c.1624G>A"	"Likely Pathogenic"



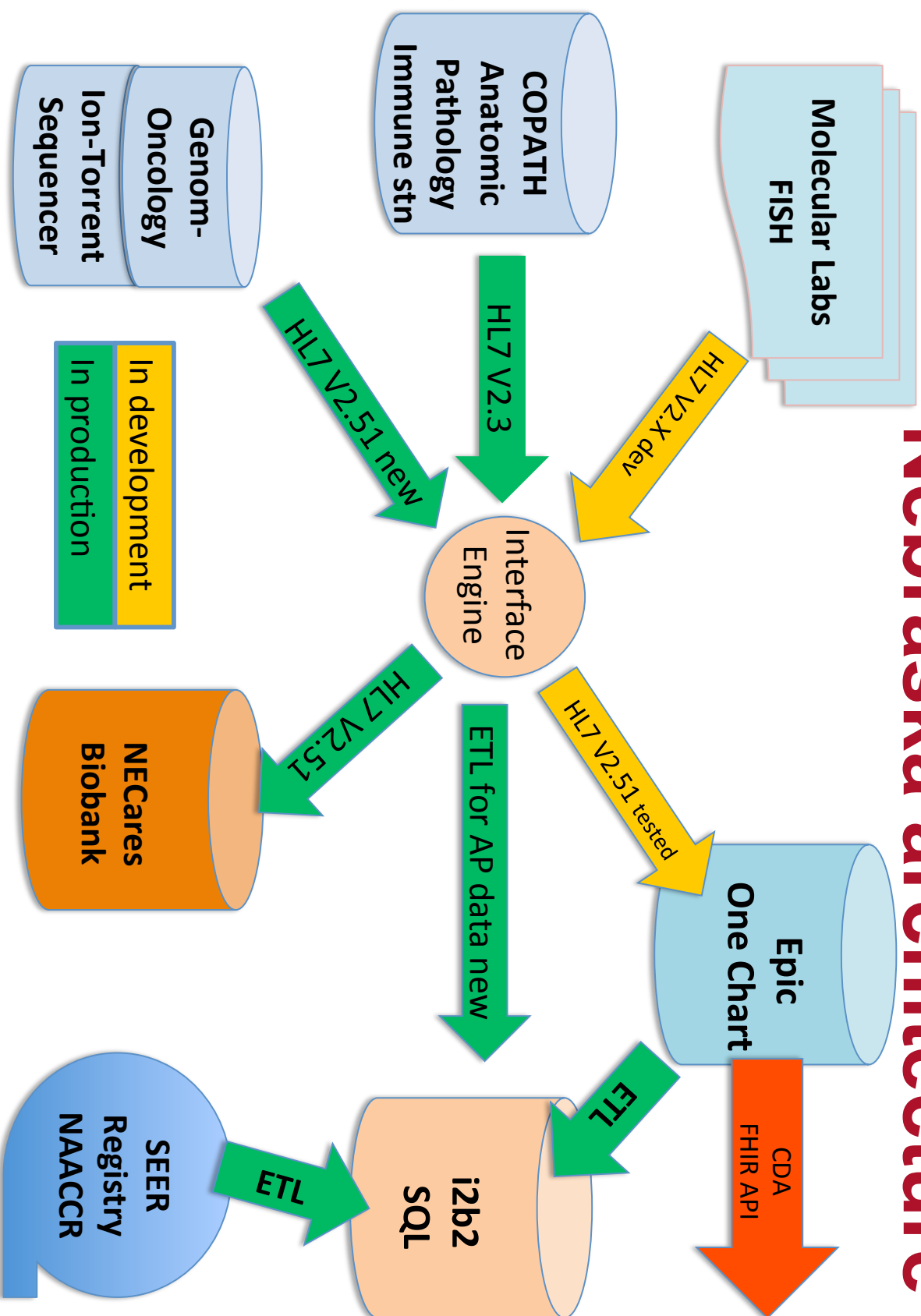
Quality

Frequency of BRAF mutations by disorder

Frequency	HGVs	Observable	Pathogenicity	Diagnosis
99	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Other"
95	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Colorectal Cancer"
59	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Non-Small Cell Lung Cancer"
15	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Tumor of Unknown Origin"
15	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Melanoma"
12	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"GIST"
12	"BRAF null:V600E null:c.1799T>A"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Pathogenic"	"Colorectal Cancer"
8	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Breast Cancer"
6	"BRAF null:V600E null:c.1799T>A"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Pathogenic"	"Melanoma"
4	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	(empty)
3	"BRAF NP_004324:V600E NIM_004333.c.1799T>A"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Pathogenic"	"Melanoma"
3	"BRAF NIM_004333.4:c.(=)"	"BRAF sequence variant identified in excised malignant neoplasm (observable entity)"	"Normal"	"Glioma"



Structured pathology reporting: Nebraska architecture





Binding Histopathology and Molecular pathology data

1. Structured and semantically encoded data
 1. Histopathology
 1. Physical morphologies
 2. Immunohistochemistry
 2. Molecular pathology
 1. Gene sequences
 2. Pathogenicity
2. Ability to bind the two



Data in action: Existing study


1. Identify all patients diagnosed with colorectal cancer between 2013 – 2016
2. Metastatic disease at diagnosis or developed in time period
3. Identify all patients with biomarker testing
 1. Microsatellite Instability (MLH1, MSH2, MSH6, PMS2)
 2. BRAF and *RAS genes
4. Which patients received targeted therapies per guidelines
 1. BRAF inhibitors
 2. EGFR inhibitors
 3. Immunotherapy
5. Pre-2015 requires manual chart review
6. 2015 – current can be computed



Stage IV Colorectal Cancer Treatments

Treatment	N	BRAF V600E +	*RAS Variant s	MicroSat Stability	Compliance
Dabrafenib	1	1	0	0	1
Cetuximab	3	0	0	0	3
Panitumumab	14	1	2	5	11
Nivolumab	1	0	0	1	1
Regorafenib	12	0	8	6	12
Totals	31	2	10	11	28





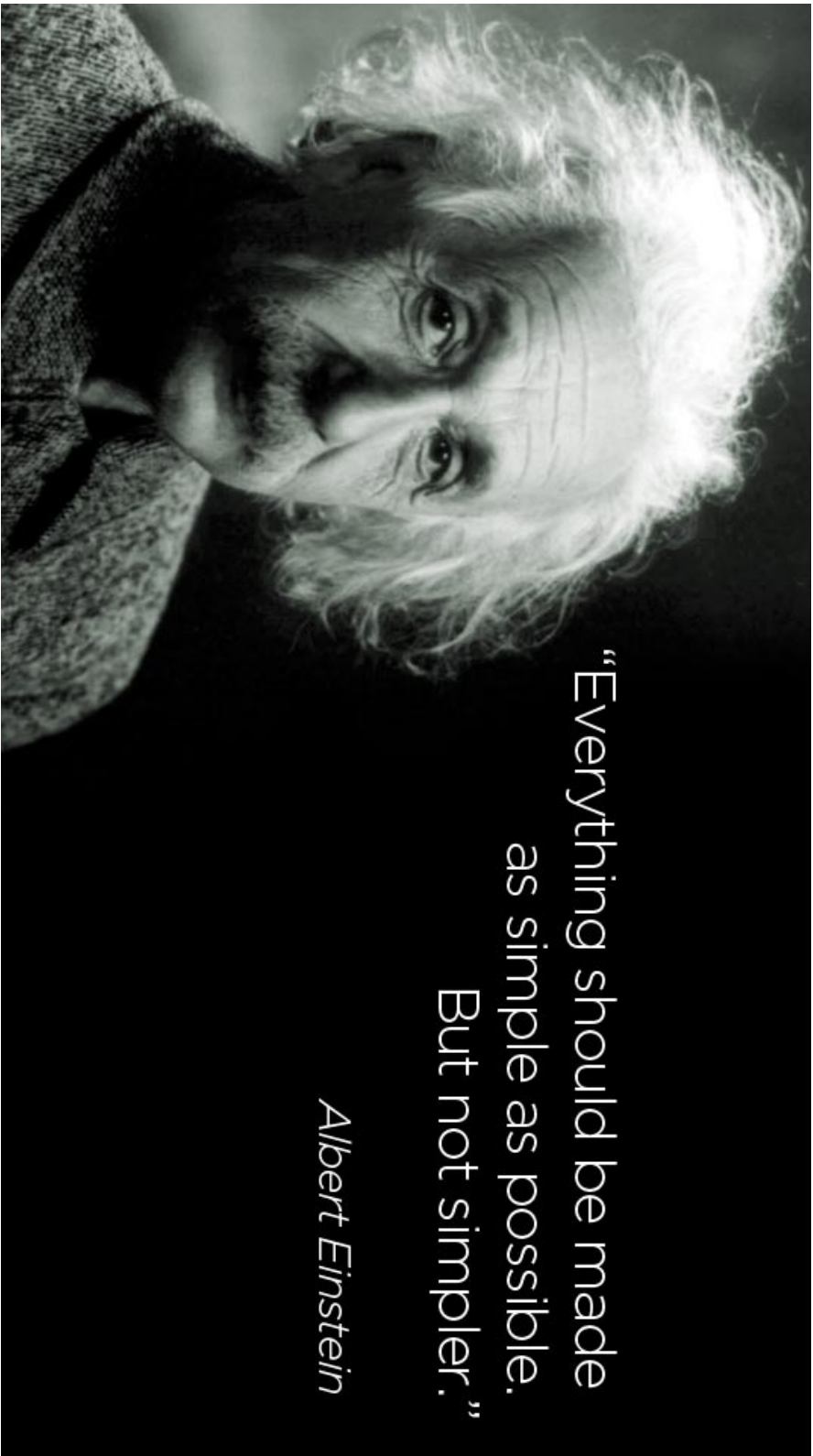
Existing Gaps and Future Work – Just some of many

1. Discrete outcome indication from clinical service
2. Additional encoded data
 1. Organ systems
 2. Support of additional genetic tests
 1. In situ hybridization (FISH)
3. Goal: Rapid identification of patients “like mine” (Phenotype)
 1. Point of care decision aid
 2. What worked well for this phenotypical patient
 3. What did not work for this phenotype
4. Clinical trial matching with high fidelity



“Everything should be made
as simple as possible.
But not simpler.”

Albert Einstein



Questions



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